

<p><b>Date of entry:</b></p>	<p>22-10-22</p>
<p><b>What have you done on your project this week?</b></p>	<p><b>AIM – to compare the OxyR regulons across different proteobacterial species</b>  Searched PubMed for papers on OxyR:  Search term “OxyR” returned 839 hits (22-10-22), while limiting to just one species “OxyR Escherichia coli” returned 412 results (some of these are still not specific to the OxyR regulon in <i>E. coli</i> – e.g. some results about <i>Franciscella</i>, <i>Paraburkholderia</i>, etc.)</p> <p>Read and critically analysed 10 of the most relevant papers looking at OxyR-regulated genes in <i>E. coli</i> (see PRISMAdiag.jpg and Endnote library for details/references)</p> <p>→Reading these papers has helped me understand which genes are regulated by OxyR in <i>E. coli</i> (and how this was demonstrated experimentally)</p>
<p><b>What have you found difficult? (How do you intend to ameliorate this? How can you grow? Can you create a bullet point for your CV from this?)</b></p>	<p>Didn’t understand the chip-exo technique used in Seo <i>et al</i> 2015 initially when I read the paper (the method used and how the data are analysed).</p> <p>Found a paper in Curr Protoc Mol Bio (Rhee and Pugh 2013) explaining the method and this helped a little.  I also found a workflow diagram made by the company AbCam (<a href="https://www.abcam.com/epigenetics/chip-20-guide-to-advanced-chromatin-immunoprecipitation-techniques">https://www.abcam.com/epigenetics/chip-20-guide-to-advanced-chromatin-immunoprecipitation-techniques</a>) illustrating the steps in ChIP-exo, which helped even more.</p> <p>CV bullet point – able to understand complex molecular biology protocols (such as ChIP-exo) and interpret the results</p>
<p><b>What has been a success?</b></p>	<p>Found a database of microarray experiments from <i>E. coli</i> (it has info on different regulons, including OxyR) - GenExpDB: <a href="https://genexpdb.okstate.edu/">https://genexpdb.okstate.edu/</a>. Looking through these data should help me understand the expression of OxyR-regulated genes in <i>E. coli</i> under different conditions.</p>
<p><b>What files/data have you produced? (are they stored securely and labelled clearly?)</b></p>	<p>Edited thesis intro: saved file with edits as thesis-intro-draft-01_Oct22_MF.docx – in “Documents/Project-Files/Drafts”</p>

	<p>Endnote library file (Project_references.enl) updated with new papers read  started to make figure 1 – model of OxyR regulon (draft) – saved as “Figure1-DRAFT1.jpg” in “Documents/Project-Files/Figures”</p>
<p><b>What is the objective for next week?</b></p>	<p>Analyse OxyR regulon in <i>E. coli</i> using the GenExpDB – and by looking for other databases (next-gen sequencing data, maybe?)</p> <p>Start looking for microarray or next-gen seq from other species (probably start with NCBI databases)</p> <p>Continue reading published papers looking at the OxyR regulons in different proteobacteria and analysing their data</p>